

Note

Changes in Sugar Content and Antioxidant Activity of *Allium* Vegetables by Salinity-stress

Mika ARAKAKI^{1,2}, Makoto TAKAHASHI², Md. Amzad HOSSAIN² and Koji WADA^{2*}

¹United Graduate School of Agricultural Science, Kagoshima University, 1-21-24 Korimoto, Kagoshima 890-8580, Japan

²Faculty of Agriculture, University of the Ryukyus, 1 Senbaru, Nishihara, Okinawa 903-0213, Japan

Received December 25, 2013 ; Accepted January 24, 2014

Three *Allium* vegetables, Welsh onion from Okinawa, and Wakegi from Okinawa and Nagasaki were subjected to salinity-stress using seawater. The K^+/Na^+ ratio of the three *Allium* vegetables clearly decreased; however, there was no significant effect on growth parameters. Of the three *Allium* vegetables, 10% seawater treatment of Welsh onion and Wakegi from Nagasaki clearly enhanced both the sugar content and antioxidant activity. Therefore, seawater treatment may be potentially useful for the development of value-added *Allium* vegetables, enhancing the palatability and food functionality.

Keywords: *Allium* vegetable, salinity-stress, seawater, sugar content, antioxidant activity

Introduction

Salinity is a major environmental factor limiting crop growth and productivity (Rhodales and Loveday, 1990). During the onset and development of salinity-stress within a crop plant, major processes such as photosynthesis, protein synthesis, and energy and lipid metabolism are affected, leading to qualitative and yield losses in most crops (Hasegawa *et al.*, 2000; Hagemann and Erdmann, 1997; Hayashi and Murata 1998). However, there are several crops with an inherent capacity to withstand salinity-stress, which allows for stable vegetable production and significantly contributes to palatability and food functionality (Sato *et al.*, 2006). Thus, there have been several reports on the application of salinity-stress for improving the quality of vegetables such as tomato fruits (Auerswald *et al.*, 1999), spinach (Makabe and Tanii, 2008), and strawberry (Keutgen and Pawelzik, 2000).

Welsh onion (*Allium fistulosum* L.), a member of the *Allium* family, is a popular vegetable in Asian countries and a reputed good source of phenol compounds with antioxidant activity, leading to beneficial medicinal effects (Helen *et al.*, 2000; Wang *et al.*, 2006). Hence, we focused our attention on exploiting salinity-stress treatment for the development of value-added Welsh onion with augmented

palatability and food functionality. There are relatively few studies on the salt tolerance of Welsh onion. In this study, we investigate the effects of artificial salinity-stress on the growth, relative chlorophyll content, and K^+/Na^+ ratio of *Allium* vegetables: Welsh onion and Wakegi (*Allium x wakegi* A.), and assess the effect of salinity-stress on sugar content, total phenol content, and antioxidant activity.

Materials and Methods

Cultivation The bulbs of three types of Japanese *Allium* cultivars, green-leafy Welsh onion (Okinawa) and two different cultivation lines of Wakegi from Okinawa and Nagasaki (Nagasaki-Wakegi) were used in this study. The upper end of each bulb was cut to adjust the overall length to 5 cm before cultivation. Each bulb type was randomly divided into four test groups of 15 plants each. One group was treated with distilled water (control), and the other three groups were treated with different concentrations of seawater (2% seawater: SW 1; 10% seawater: SW 2; 100% seawater: SW 3) to induce salinity-stress. Each type of bulb was cultivated in a Wagner pot (1/5000 a) with three plants per pot (total of five pots per test group; plant cultivation was initiated on November 19, 2012). After two weeks of cultivation, the plant leaves were thoroughly sprayed

*To whom correspondence should be addressed.

E-mail: kojiwada@agr.u-ryukyu.ac.jp

with seawater at 5 mL of seawater per plant, and the same treatment was repeatedly conducted four times in total every 5 days. The mineral content of the 100% seawater used in this study was as follows (mg/100 mL): Na⁺: 942.7; K⁺: 28.68; Ca²⁺: 38.5; Mg²⁺: 140.7. Five days after the last treatment, the edible portions of each plant were used in the experiment. In the case of growth parameter measurement, the plants (n = 15) were individually analyzed from five pots. Next, three pots of plants were randomly selected from five pots and promptly freeze-dried for mineral analysis. The remainder was collectively analyzed for sugar content, total phenol content, and antioxidant activity measurements, respectively.

Growth parameters and K⁺/Na⁺ ratio measurement The growth parameters (plant height, leaf blade diameter, total weight, number of leaves, number of tillers, number of broken leaves, number of dead leaves, and soil and plant analyzer development (SPAD) value as relative chlorophyll content) were evaluated. The SPAD value was measured using a chlorophyll meter (SPAD-502, Minolta Co., Osaka, Japan) and recorded as the mean of ten measurements for each individual leaf (Cho *et al.*, 2007). The Na⁺ and K⁺ contents of the freeze-dried leaves were determined using inductively coupled plasma/atomic emission spectroscopy (Zeinera *et al.*, 2005) at respective wavelengths of 330.232 nm and 797.395 nm, on an ICPE-9000 spectrometer (Shimadzu Co., Kyoto, Japan). The levels were expressed in terms of the K⁺/Na⁺ ratio.

Sugar content analysis The free sugar content was determined according to the method described by Miyagi *et al.* (2011). Briefly, a 5 mm section of each plant (10 g) was refluxed in 100 mL of 85% ethanol (80°C, 60 min). The ethanol extract was filtered and adjusted to 200 mL with 85% ethanol. A 20-mL aliquot of the diluted extract was evaporated to dryness *in vacuo*, and the residue was dissolved in 50 mL of distilled water. An aliquot of this solution was analyzed for the content of reducing (glucose and fructose) and non-reducing (sucrose) sugars using high-performance liquid chromatography on an anion-exchange column (250 mm × 2.0 mm) coupled with an electrochemical detector (Dionex Co., Osaka, Japan). The total sugars were also calculated as the total quantity of each sugar.

Total phenol content analysis Samples for the evaluation of total phenol content were prepared based on the method described by Žitňanova *et al.* (2006). Briefly, a 5 mm section of each plant (10 g) and distilled water (90 g) were mixed and homogenized in a high-speed blender. The obtained homogenates were centrifuged at 4500 × g for 5 min and the supernatants were assayed for total phenol content of the *Allium* vegetable samples. The total phenol content of the samples was evaluated using the Folin-Ciocalteu method of Sato *et al.* (2010). The total phenol content was calculated from a calibration curve of gallic acid (10 – 150 µg/mL) and is expressed as milligrams of gallic acid equivalents (GAE) per 100-grams fresh weight (FW).

Antioxidant activity measurement Samples for the evaluation of antioxidant activity were prepared as for the above total phenol content analysis. The antioxidant activity of the samples was chemically evaluated by determination of the oxygen radical

absorbance capacity (ORAC), which has the advantage of utilizing free radical generators, and produces the biologically relevant peroxy radical (Ishimoto *et al.*, 2012). The ORAC assay was performed according to the method reported by Prior *et al.* (2003). 2,2'-Azobis(2-amidinopropane)dihydrochloride (Wako Pure Chemical Industries, Osaka, Japan) was used as a peroxy radical generator, and fluorescein (Sigma-Aldrich, St. Louis, MO, USA) and 6-hydroxy-2,3,7,8-tetramethylchroman-2-carboxylic acid (Trolox; Calbiochem, San Diego, CA, USA) were respectively used as a fluorescent probe (Naguib, 2000) and standard. The ORAC value was calculated by using a quadratic regression equation relating the Trolox or sample concentration and area under the curve (AUC) of the fluorescence, and is expressed as micromoles of Trolox equivalents (TE) per 100-grams FW.

Statistical analysis The data of growth parameters (n = 15) and K⁺/Na⁺ ratio (n = 3) of the three *Allium* vegetables are expressed as mean ± SE. In the case of sugar content, total phenol content and antioxidant activity measurements, each measurement was performed in triplicate and values are expressed as an average. The data of growth parameters and K⁺/Na⁺ ratio were analyzed using a one-way analysis of variance with Dunnett's multiple comparison test. Differences were considered significant at *p* < 0.05 or 0.01.

Results and Discussion

Effect of salinity-stress on growth parameters and K⁺/Na⁺ ratio of *Allium* vegetables Table 1 shows the growth parameters and K⁺/Na⁺ ratio of the three *Allium* vegetables subjected to salinity-stress using different concentrations of seawater. None of the growth parameters, except for SPAD value, was clearly related to the degree of salinity-stress for the three *Allium* vegetables. Generally, salinity-stress results in a clear stunting of plants. Wang *et al.* (2000) reported that a reduction in the rate of leaf surface expansion with increasing salt concentration is an immediate response of salinity-stress. Salinity-stress also results in a considerable decrease in the fresh weights of leaves, stems, and roots (Chartzoulakis and Klapaki, 2000).

Suppression of growth occurs in most plants; however, the tolerance level and rate of growth reduction under salinity-stress vary widely among different plant species or cultivar lines (Parida and Das, 2005). Herein, the SPAD values of the leaves of the two types of Wakegi cultivars increased, whereas the SPAD value of the Welsh onion decreased, similar to the general response of most plants (Khavarinejad and Chaparzadeh, 1998). Although the salinity was found to induce little growth suppression in the three *Allium* vegetables, further investigation into the effects of other seawater loading methods on growth parameters is necessary, as properties, especially plant size, form, and color, also characterize the product value of vegetables.

The K⁺/Na⁺ ratio clearly decreased with an increase of seawater concentration up to 100% in all cases. The maintenance of a high K⁺/Na⁺ ratio is closely related to salt tolerance in several plants

Table 1. Comparison of the growth parameters and K⁺/Na⁺ ratio of the three *Allium* vegetables with or without salinity-stress treatment for 40 days

<i>Allium</i> vegetable	Group	Plant height (cm)	Leaf blade diameter (mm)	Total weight (g)	Number of leaves	Number of tillers	Number of broken leaves	Number of dead leaves	SPAD value	K ⁺ /Na ⁺ ratio
Welsh onion	Control	28.29 ± 2.5	4.37 ± 0.9	8.48 ± 1.9	8.47 ± 1.6	1.80 ± 0.4	2.20 ± 1.5	2.00 ± 0.7	59.69 ± 1.3	14.41 ± 3.8
	SW 1	25.51 ± 2.7	3.43 ± 0.7*	5.79 ± 1.5*	7.53 ± 1.8	1.47 ± 0.4	1.60 ± 1.3	1.80 ± 0.8	58.45 ± 1.9	12.75 ± 0.8
	SW 2	29.23 ± 3.2	4.00 ± 0.2	6.95 ± 1.2	7.93 ± 1.9	1.60 ± 0.6	2.00 ± 0.7	1.60 ± 1.1	57.15 ± 2.2*	10.45 ± 2.7
	SW 3	27.65 ± 2.5	3.63 ± 0.2	5.82 ± 0.9*	7.00 ± 1.6	1.67 ± 0.5	2.60 ± 1.5	0.60 ± 0.5*	57.77 ± 1.7	2.53 ± 0.7**
Wakegi (Okinawa)	Control	33.67 ± 2.8	5.57 ± 0.5	15.13 ± 1.9	11.40 ± 1.1	2.00 ± 0.2	0.80 ± 1.1	3.40 ± 1.5	58.27 ± 0.4	30.36 ± 2.4
	SW 1	32.35 ± 3.6	5.93 ± 0.9	13.73 ± 2.5	10.93 ± 0.4	2.00 ± 0.2	1.00 ± 0.7	2.20 ± 0.8	60.87 ± 1.5	26.23 ± 0.5*
	SW 2	36.06 ± 4.9	5.10 ± 0.7	17.01 ± 5.0	13.07 ± 2.6	2.27 ± 0.3	0.80 ± 0.8	1.80 ± 0.8*	62.59 ± 1.1	20.06 ± 3.3**
	SW 3	33.87 ± 3.5	4.97 ± 0.6	14.91 ± 2.3	11.93 ± 1.6	2.47 ± 0.2*	1.00 ± 0.7	2.40 ± 0.9	61.02 ± 1.4	3.83 ± 0.6**
Wakegi (Nagasaki)	Control	24.63 ± 2.7	3.70 ± 0.3	9.42 ± 1.9	12.47 ± 2.2	2.40 ± 0.4	0.60 ± 0.5	0.00 ± 0.0	58.80 ± 2.5	11.996 ± 0.5
	SW 1	23.68 ± 3.9	3.37 ± 0.5	8.22 ± 2.4	11.53 ± 2.4	2.53 ± 0.3	0.40 ± 0.9	0.60 ± 0.9	57.29 ± 0.9	11.168 ± 2.0
	SW 2	21.75 ± 3.8	3.43 ± 0.7	7.16 ± 2.5	10.13 ± 2.1	2.33 ± 0.6	0.00 ± 0.0	0.60 ± 0.5	61.76 ± 1.5*	7.675 ± 2.5*
	SW 3	24.36 ± 5.9	3.50 ± 0.9	8.25 ± 2.8	10.33 ± 1.2	2.40 ± 0.4	0.20 ± 0.4	2.80 ± 1.9*	62.35 ± 2.8*	2.831 ± 0.4**

The values of growth parameters (n = 15) and K⁺/Na⁺ ratio (n = 3) are shown as mean ± SE. The seawater concentrations were 2% (SW 1), 10% (SW 2), and 100% (SW 3). A difference between the control group and each seawater treated group (SW 1, 2, and 3) is considered statistically significant when *p < 0.05 or **p < 0.01.

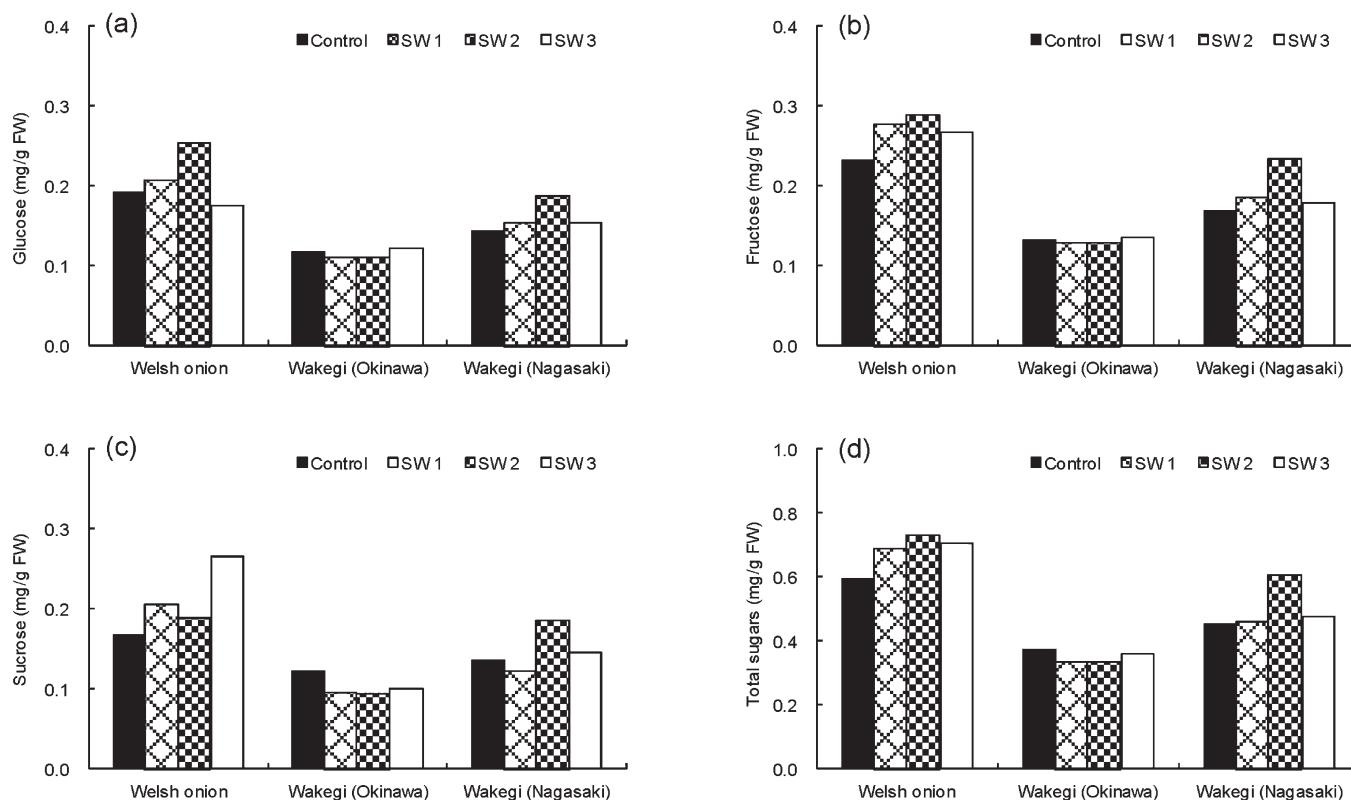


Fig. 1. Effect of salinity-stress on the contents of glucose (a), fructose (b), sucrose (c), and total sugars (d) in leaves of the three *Allium* vegetables with or without salinity-stress treatment for 40 days. Averaged values are shown. The seawater concentrations were 2% (SW 1), 10% (SW 2), and 100% (SW 3).

(Gadallah, 1999; Sobahan *et al.*, 2012). For example, salinity-stress decreases the K^+/Na^+ ratio of typical rice cultivars due in part to increased Na^+ uptake, mediated by the apoplastic pathway, in cultivars under salinity-stress (Sobahan *et al.*, 2012). Indeed, the Na^+ contents of all of the evaluated *Allium* vegetables increased with increasing seawater concentration, whereas the K^+ content was not significantly affected (data not shown). Therefore, this finding suggests that the K^+/Na^+ ratio of Welsh onion and Wakegi leaves may serve as a possible index of salinity-stress, at least in case of direct seawater-spray on leaves.

Effect of salinity-stress on sugar content, total phenol content, and antioxidant activity of Allium vegetables Plants exposed to salinity-stress in their environment undergo changes in growth parameters, as well as in the synthesis of active metabolites that control ion and water flux and support scavenging of reactive oxygen species (ROS) (Gadallah, 1999). To elucidate the effects of salinity-stress on active metabolites, the sugar and total phenol contents, as well as the antioxidant activity of Welsh onion and Wakegi were measured. Evaluation of the three plants indicated that the reducing (Fig. 1-a and Fig. 1-b) and non-reducing (Fig. 1-c) sugar contents of the Welsh onion and Wakegi from Nagasaki increased in response to salinity-stress treatment, and the total sugar content increased considerably compared to the respective controls for the 10% seawater concentration (Fig. 1-d). In contrast, the total sugar content was lowered relative to the control only in the case of Wakegi from Okinawa (Fig. 1-d). Salinity-stress increases the reducing and non-

reducing sugars in a number of plants due to activation of their osmoprotective, osmotic adjustment, carbon storage, and radical scavenging functions (Kerepesi and Galiba, 2000). Therefore, it was suggested that salt tolerance mechanisms of Wakegi from Okinawa might differ from those of Welsh onion and Wakegi from Nagasaki. Miyagi *et al.* (2011) conducted sensory evaluation of Welsh onion, and demonstrated a strong positive correlation between free sugar content and palatability. In the case of Welsh onion and Wakegi from Nagasaki, salinity treatment with a seawater concentration of 10% effectively produced a higher content of sugars compared to the control plants not subjected to the treatment, which might lead to good palatability. Figure 2 shows the respective total phenolic contents (a) and ORAC values (b) of the three *Allium* vegetables in response to salinity-stress. Generally, salinity-stress leads to the formation of ROS in plants, which produces a number of antioxidative phenolic compounds that protect against the potentially cytotoxic activated oxygen species (Gadallah, 1999). Of the two Wakegi cultivars, there was a clear increase in the total phenol content and ORAC value of Wakegi from Nagasaki compared to the control in response to salinity-stress of 2% and 10% seawater, whereas these values did not change markedly in Wakegi from Okinawa. Interestingly, the ORAC value of the Welsh onion was increased notably due to salinity-stress, despite the considerable decrease of its total phenol content.

Accumulation of antioxidants such as phenol compounds and ascorbic acid (Ishikawa and Shigeoka, 2008), glutathione (Gadallah,

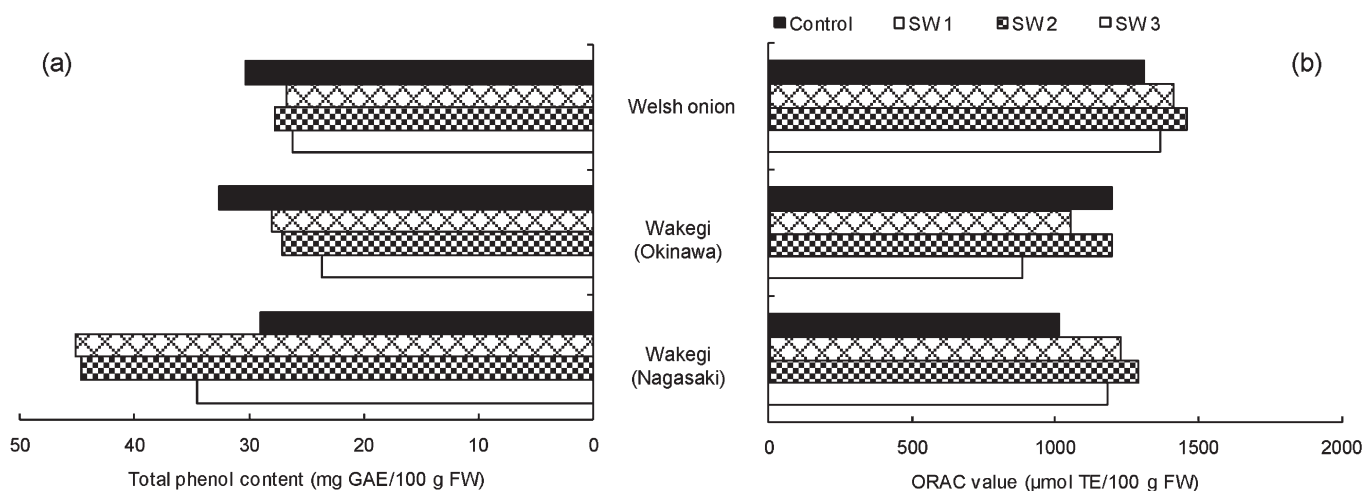


Fig. 2. Total phenolic content (a) and Oxygen Radical Absorbance Capacity (ORAC) values (b) of the three *Allium* vegetables with or without salinity-stress treatment for 40 days.

Averaged values are shown. The seawater concentrations were 2% (SW 1), 10% (SW 2), and 100% (SW 3).

1999), as well as volatile compounds (Zuo *et al.*, 2012) under salinity-stress has been noted in several plants. Indeed, Welsh onion possesses antioxidant compounds such as quercetin and kaempferol as main phenol compounds (Wang *et al.*, 2006), ascorbic acid (Kähkönen and Heinonen, 2003), and volatile sulfur compounds (Wu *et al.*, 2001). Thus, it is possible that the content of other antioxidant compounds besides phenol compounds may be increased in the Welsh onion, thereby increasing the antioxidant activity under salinity-stress. From a functional food perspective, seawater treatment is thus deemed highly advantageous in the case of Welsh onion and Wakegi from Nagasaki. It was found herein that salinity treatment at a seawater concentration of 10% clearly enhanced both the sugar content and antioxidant activity of these species. However, the active component in Welsh onion and Wakegi from Nagasaki that is responsible for the antioxidant activity was not elucidated, and further studies are therefore required.

In conclusion, salinity-stress using seawater may augment the sugar content, antioxidant activity and other growth parameters, allowing for the development of value-added *Allium* vegetables with enhanced palatability and food functionality.

References

- Auerswald, H., Schwarz, D., Kornelson, C., Krumbein, A., and Bruckner, B. (1999). Sensory analysis, sugar and acid content of tomato at different EC values of nutrient solution. *Sci. Hort.*, **82**, 227-242.
- Chartzoulakis, K. and Klapaki, G. (2000). Response of two green house pepper hybrids of NaCl salinity during different growth stages. *Sci. Hort.*, **86**, 247-260.
- Cho, Y.Y., Oh, S., Oh, M.M., and Son, J.E. (2007). Estimation of individual leaf area, fresh weight, and dry weight of hydroponically grown cucumbers (*Cucumis sativus* L.) using leaf length, width, and SPAD value. *Sci. Hort.*, **111**, 330-334.
- Gadallah, M.A.A. (1999). Effects of proline and glycinebetaine on *Vicia faba* response to salt stress. *Biol. Plant.*, **42**, 249-257.
- Hagemann, M. and Erdmann, N. (1997). Environmental stresses. In "Cyanobacterial nitrogen metabolism and environmental biotechnology." ed. by A.K. Rai. Springer-Verlag, New Delhi, pp. 156-221.
- Hasegawa, P.M., Bressan, R.A., Zhu, J.K., and Bohnert, H.J. (2000). Plant cellular and molecular responses to high salinity. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, **51**, 463-499.
- Hayashi, H. and Murata, N. (1998). Genetically engineered enhancement of salt tolerance in higher plants. In "Stress responses of photosynthetic organisms." eds. by K. Sato and N. Murata. Elsevier, Amsterdam, pp. 133-148.
- Helen, A., Krishnakumar, K., Vijayammal, P.L., and Augusti, K.T. (2000). Antioxidant effect of onion oil (*Allium cepa*, Linn) on the damages induced by nicotine in rats as compared to alpha-tocopherol. *Toxicol. Letters*, **116**, 61-68.
- Ishikawa, T. and Shigeoka, S. (2008). Recent advances in ascorbate biosynthesis and the physiological significance of ascorbate peroxidase in photosynthesizing organisms. *Biosci. Biotechnol. Biochem.*, **72**, 1143-1154.
- Ishimoto, H., Tai, A., Yoshimura, M., Amakura, Y., Yoshida, T., Hatano, T., and Ito, H. (2012). Antioxidative properties of functional polyphenols and their metabolites assessed by an ORAC assay. *Biosci. Biotechnol. Biochem.*, **76**, 395-399.
- Kähkönen, M.P. and Heinonen, M. (2003). Antioxidant activity of anthocyanins and their aglycons. *J. Agric. Food Chem.*, **51**, 628-633.
- Kerepesi, I. and Galiba, G. (2000). Osmotic and salt stress-induced alteration in soluble carbohydrate content in wheat seedlings. *Crop Sci.*, **40**, 482-487.
- Keutgen, A.J. and Pawelzik, E. (2008). Quality and nutritional value of strawberry fruit under long term salt stress. *Food Chem.*, **107**, 1413-1420.
- Khavarinejad, R.A. and Chaparzadeh, N. (1998). The effects of NaCl and CaCl₂ on photosynthesis and growth of alfalfa plants. *Photosynthetica*, **35**, 461-466.
- Makabe, Y. and Tanii, J. (2008). Use development by scattering salt in

- vegetable farms (II) - cultivation of spinach -. *Bull. Soc. Sea Water Sci. Jpn.*, **62**, 191-193 (in Japanese).
- Miyagi, A., Yasuda, M., Hisaka, H., Motoori, S., and Wako, T. (2011). Relationship between sensory test, and chemical and/or physical properties of Japanese bunching onion. *Hort. Res.*, **10**, 101-107 (in Japanese).
- Naguib, Y.M. (2000). A fluorometric method for measurement of oxygen radical-scavenging activity of water-soluble antioxidants. *Anal. Biochem.*, **284**, 93-98.
- Parida, A.K. and Das, A.B. (2005). Salt tolerance and salinity effects on plants: a review. *Ecotox. Environ. Safe.*, **60**, 324-349.
- Prior, R.L., Hoang, H., Gu, L., Wu, X., Bacchiocca, M., Howard, L., Woodill, M.H., Huang, D., Ou, B., and Jacob, R. (2003). Assays for hydrophilic and lipophilic antioxidant capacity (oxygen radical absorbance capacity (ORAC_{FL})) of plasma and other biological and food samples. *J. Agric. Food Chem.*, **5**, 3273-3279.
- Rhoades, J.D. and Loveday, J. (1990). Salinity in irrigated agriculture. In "American Society of Civil Engineers." eds. by B.A. Steward and D.R. Nielsen. American Society of Agronomists, Madison, pp. 1089-1142.
- Sato, A., Watanabe, J., Goto, M., and Takano, Y.I. (2010). Evaluation for anti-oxidant activities of *Prunus* sp. using oxygen radical absorbance capacity. *Nippon Shokuhin Kagaku Kogaku Kaishi*, **57**, 44-48 (in Japanese).
- Sato, S., Sakaguchi, S., Furukawa, H., and Ikeda, H. (2006). Effects of NaCl application to hydroponic nutrient solution on fruit characteristic of tomato (*Lycopersicon esculentum* Mill.). *Sci. Hort.*, **109**, 248-253.
- Sobahan, M.A., Akter, N., Ohno, M., Okuma, E., Hirai, Y., Mori, I.C., Nakamura, Y., and Murata, Y. (2012). Effects of exogenous proline and glycinebetaine on the salt tolerance of rice cultivars. *Biosci. Biotechnol. Biochem.*, **76**, 1568-1570.
- Wang, B.S., Lin, S.S., Hsiao, W.C., Fan, J.J., Fuh, L.F., and Duh, P.D. (2006). Protective effects of an aqueous extract of Welsh onion green leaves on oxidative damage of reactive oxygen and nitrogen species. *Food Chem.*, **98**, 149-157.
- Wang, Y. and Nil, N. (2000). Changes in chlorophyll, ribulose biphosphate carboxylase-oxygenase, glycine betaine content, photosynthesis and transpiration in *Amaranthus tricolor* leaves during salt stress. *J. Hortic. Sci. Biotechnol.*, **75**, 623-627.
- Wu, C.C., Sheen, L.Y., Chen, W.H., Tsai, S.J., and Lii, C.K. (2001). Effect of organosulfur compounds from garlic oil on the antioxidation system in rat liver and red blood cells. *Food Chem. Toxicol.*, **39**, 563-569.
- Zeinera, M., Steffana, I., and Cindric, I.J. (2005). Determination of trace elements in olive oil by ICP-AES and ETA-AAS: A pilot study on the geographical characterization. *Microchem. J.*, **81**, 171-176.
- Žitňanova, I., Ranostajova, S., Sobotova, H., Demelova, D., Pechaň, I., and Ďuračková, Z. (2006). Antioxidative activity of selected fruits and vegetables. *Biologia*, **61**, 279-284.
- Zuo, Z.J., Zhu, Y.R., Bai, Y.L., and Wang, Y. (2012). Volatile communication between *Chlamydomonas reinhardtii* cells under salt stress. *Biochem. Syst. Ecol.*, **40**, 19-24.